

# Maize GOLDEN2-LIKE proteins enhance drought tolerance in rice by promoting stomatal closure

Xia Li <sup>1,†</sup> Jing Li <sup>1,†</sup> Shaobo Wei <sup>1</sup> Yuan Gao <sup>1</sup> Hongcui Pei <sup>1</sup> Rudan Geng <sup>1</sup> Zefu Lu <sup>1</sup>  
Peng Wang <sup>2</sup> and Wenbin Zhou <sup>1,\*</sup>

1 Institute of Crop Sciences, Chinese Academy of Agricultural Sciences, Beijing 100081, China

2 CAS Center for Excellence in Molecular Plant Sciences, Institute of Plant Physiology and Ecology, Chinese Academy of Sciences, Shanghai 200032, China

\*Author for correspondence: zhouwenbin@caas.cn

<sup>†</sup>These authors contributed equally.

The author responsible for distribution of materials integral to the findings presented in this article in accordance with the policy described in the Instructions for Authors (<https://academic.oup.com/plphys/pages/General-Instructions>) is Wenbin Zhou (zhouwenbin@caas.cn).

## Abstract

Drought has become one of the most severe abiotic stresses experienced in agricultural production across the world. Plants respond to water deficit via stomatal movements in the leaves, which are mainly regulated by abscisic acid (ABA). A previous study from our lab showed that constitutive expression of maize (*Zea mays* L.) GOLDEN2-LIKE (GLK) transcription factors in rice (*Oryza sativa* L.) can improve stomatal conductance and plant photosynthetic capacity under field conditions. In the present study, we uncovered a function of ZmGLK regulation of stomatal movement in rice during drought stress. We found that elevated drought tolerance in rice plants overexpressing *ZmGLK1* or *GOLDEN2* (*ZmG2*) was conferred by rapid ABA-mediated stomatal closure. Comparative analysis of RNA-sequencing (RNA-seq) data from the rice leaves and DNA affinity purification sequencing (DAP-seq) results obtained in vitro revealed that ZmGLKs played roles in regulating ABA-related and stress-responsive pathways. Four upregulated genes closely functioning in abiotic stress tolerance with strong binding peaks in the DAP-seq data were identified as putative target genes of ZmGLK1 and ZmG2 in rice. These results demonstrated that maize GLKs play an important role in regulating stomatal movements to coordinate photosynthesis and stress tolerance. This trait is a valuable target for breeding drought-tolerant crop plants without compromising photosynthetic capacity.

Received August 24, 2023. Accepted September 29, 2023. Advance access publication October 18, 2023

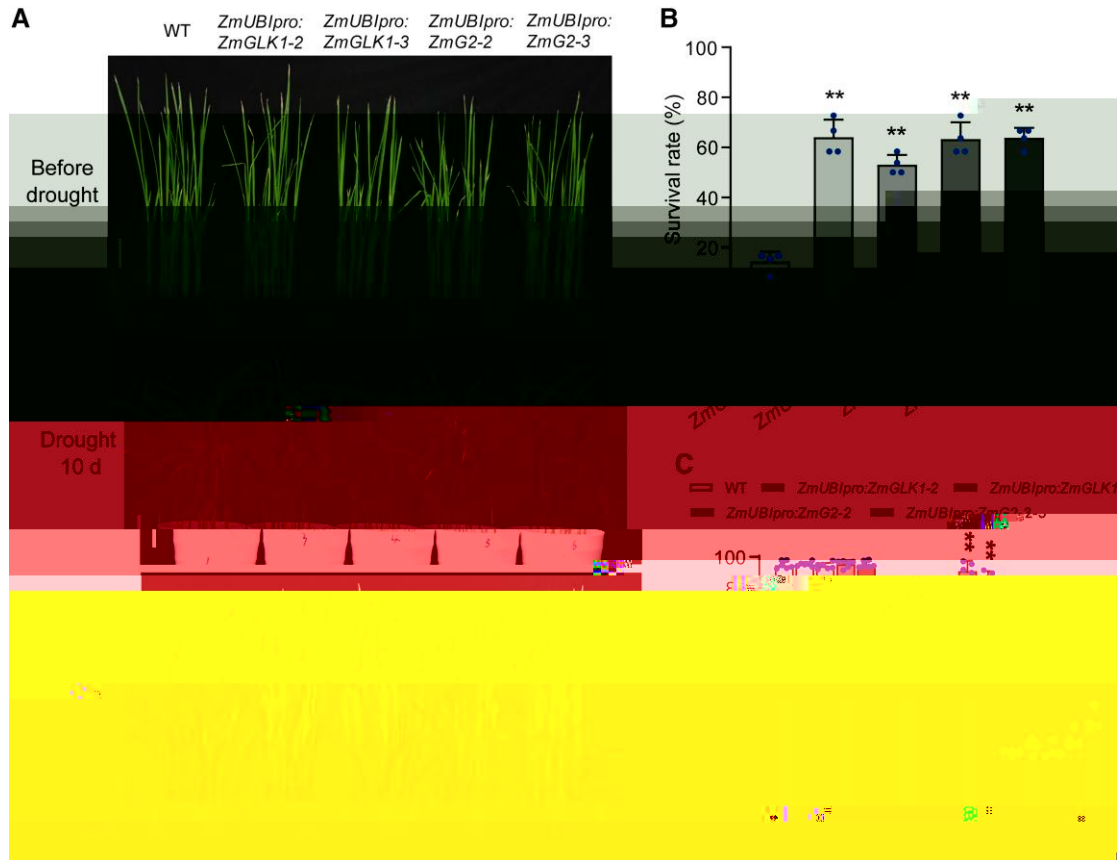
© The Author(s) 2023. Published by Oxford University Press on behalf of American Society of Plant Biologists.

This article is distributed under the terms of the Creative Commons Attribution License (<https://creativecommons.org/licenses/by/4.0/>)

Open Access

deficit or are exposed to other environmental stimuli (e.g. low light intensity, low air humidity, high CO<sub>2</sub> levels, and pathogens), stomata are rapidly closed, especially in angiosperms (Sierla et al. 2018). This dynamic movement is driven by turgor pressure changes in the guard cells, as a result of the activation of anion channels and the inhibition of inward-rectifying K<sup>+</sup> channels, which encoded by *K<sup>+</sup> CHANNEL IN ARABIDOPSIS THALIANA* (*KAT*) and *ARABIDOPSIS K<sup>+</sup> TRANSPORTER* (*AKT*) genes (Kim et al. 2010). The efflux of anions and small metabolites, including Cl<sup>-</sup>, NO<sub>3</sub><sup>-</sup>, and malate, causes membrane depolarization to activate the outward-rectifying K<sup>+</sup> channel and facilitates K<sup>+</sup> efflux, further reducing turgor pressure inside the guard cells and leading to the stomatal closure (Pandey et al. 2007). Under water-deficit conditions, the phytohormone abscisic acid (ABA) plays as the primary regulator of stomatal movement to prevent water loss, in which endogenous ABA levels are controlled by a precise balance between biosynthesis and catabolism, which also influenced by transport and conjugation process (Kushiro et al. 2004; Hsu et al. 2021). ABA is initially synthesized from C<sub>40</sub> carotenoids to form xanthophylls (e.g. 9-*cis*-violaxanthin and 9-*cis*-neoxanthin); a C<sub>15</sub> intermediate, xanthoxin, is formed in the plastids via oxidative cleavage catalyzed by 9-*cis*-epoxycarotenoid dioxygenase (NCED). Xanthoxin is then exported to the cytosol and converted to ABA through a 2-step reaction via short-chain dehydrogenase/reductase 1 (SDR1/ABA2) and Arabidopsis aldehyde oxidase 3 (AAO3) (Seo and Koshiba 2002; Xiong and Zhu 2003).

Transcription factors (TFs) are crucial regulators of many biological processes, including responses to environmental signals and hormone regulation. These regulatory functions are accomplished through binding to specific *cis*-elements in the promoter regions of target genes (Todaka et al. 2012). Numerous abiotic stress-responsive TFs have been identified in plants; for instance, WRKY, MYB, and DREB/CBF TFs have all been reported as key regulators of plant stress responses (Manna et al. 2021). GOLDEN2-LIKE (GLK) TFs generally act as transcriptional activators of chloroplast development and biogenesis (Rossini et al. 2001; Wang et al. 2013) and play important roles in regulating nuclear photosynthesis-related genes (Chen et al. 2016). In maize (*Zea mays* L.), 2 *GLK* genes, *ZmGLK1* and *GOLDEN2* (*ZmG2*), have shown differential expression patterns between mesophyll cells and the bundle sheath (Hall et al. 1998; Chang et al. 2012). Ectopic overexpression of maize *GLK* genes in rice induces chloroplast development in bundle sheath cells and activates intracellular plasmodesmatal con



**Figure 1.** Overexpression of *ZmGLK1* and *ZmG2* in rice increased drought tolerance. **A)** Phenotypes of WT, *ZmUBI<sub>pro</sub>:ZmGLK1*, and *ZmUBI<sub>pro</sub>:ZmG2* rice plants during drought stress. Three-week-old WT, *ZmUBI<sub>pro</sub>:ZmGLK1*, and *ZmUBI<sub>pro</sub>:ZmG2* rice seedlings grown in soil were drought stressed by withholding water for 10 d and then watered for a 6-d recovery period. The upper, middle, and lower panels show representative plants before drought stress, after 10 d of drought stress, and after the 6-d recovery, respectively. Scale bar: 2 cm. **B)** Survival rates of WT, *ZmUBI<sub>pro</sub>:ZmGLK1*, and *ZmUBI<sub>pro</sub>:ZmG2* rice plants after 10 d of drought stress followed by 6 d of recovery. Data are presented as the mean  $\pm$  SD from 4 biological replicates. **C)** The RWC of WT, *ZmUBI<sub>pro</sub>:ZmGLK1*, and *ZmUBI<sub>pro</sub>:ZmG2* rice leaves after 0, 7, and 10 d of drought stress. Data are presented as the mean  $\pm$  SD

10 d of PEG treatment (Supplemental Fig. S1B). We also monitored changes of RWC in rice seedling during PEG treatment. The results showed that the transgenic plants retained significantly higher RWC compared to the WT. Specifically, RWC values were 11.4% to 12.1% and 29.5% to 29.7% higher in *ZmUBI<sub>pro</sub>:ZmGLK1* and *ZmUBI<sub>pro</sub>:ZmG2* rice plants, respectively, compared with the WT (Supplemental Fig. S1C). These results together indicated that overexpression of *ZmGLK1* and *ZmG2* in rice significantly improve the tolerance to drought and osmotic stress.

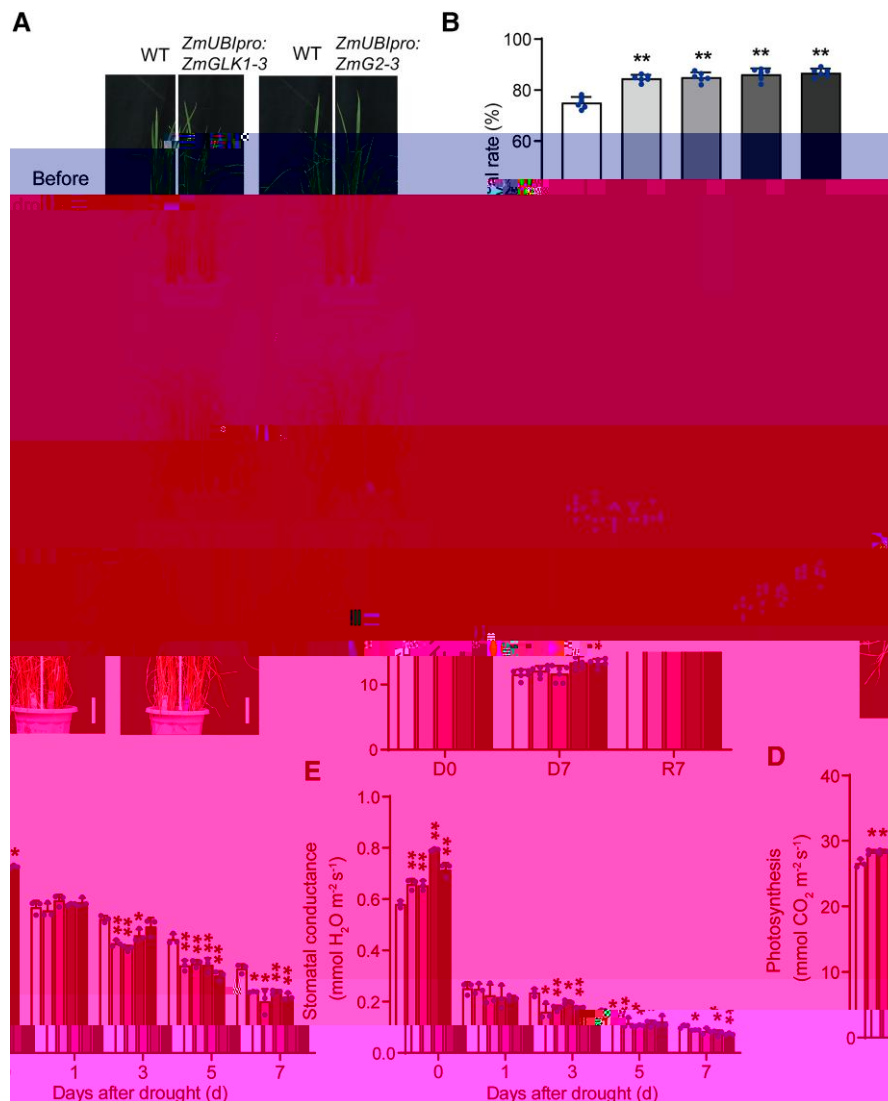
### ZmGLK1 and ZmG2 triggered rapid stomatal closure in drought-stressed rice plants

To further investigate the physiological mechanism underlying the elevated drought tolerance conferred by *ZmGLK1* and *ZmG2*, we evaluated the effects of drought treatment on stomatal traits of rice seedlings grown in the pots in the growth chamber, since stomata are the main channels for gas exchange and water respiration in plants, serving as the dominant limitation to photosynthesis under drought. We therefore first measured

stomatal conductance and photosynthetic-related parameters under control conditions using a LICOR-6400XT portable photosynthesis system. The results revealed significantly higher stomatal conductance in *ZmUBI<sub>pro</sub>:ZmGLK1* and *ZmUBI<sub>pro</sub>:ZmG2* rice seedlings (0.118–0.139 and 0.126–0.131, respectively) compared with the WT (0.083) under control condition; while the transgenic plants also performed higher photosynthesis rates, intercellular CO<sub>2</sub> concentrations (*C<sub>i</sub>*), and transpiration rates (Supplemental Fig. S2), as the plants grown in the field (Li et al. 2020). In contrast, after 7 d of drought treatment, *ZmUBI<sub>pro</sub>:ZmGLK1* and *ZmUBI<sub>pro</sub>:ZmG2* rice plants displayed sharply decrease in stomatal conductance (0.062–0.073 and 0.054–0.059, respectively), whereas that of WT remained relatively stable under drought conditions (0.087; Supplemental Fig. S2B). The photosynthesis rates, *C<sub>i</sub>*, and transpiration rates showed corresponding declines in *ZmUBI<sub>pro</sub>:ZmGLK1* and *ZmUBI<sub>pro</sub>:ZmG2* rice plants during water deprivation (Supplemental Fig. S2, A, C, and D).

We next compared the stomatal traits between WT and *ZmUBI<sub>pro</sub>:ZmGLK1* or *ZmUBI<sub>pro</sub>:ZmG2* rice plants under





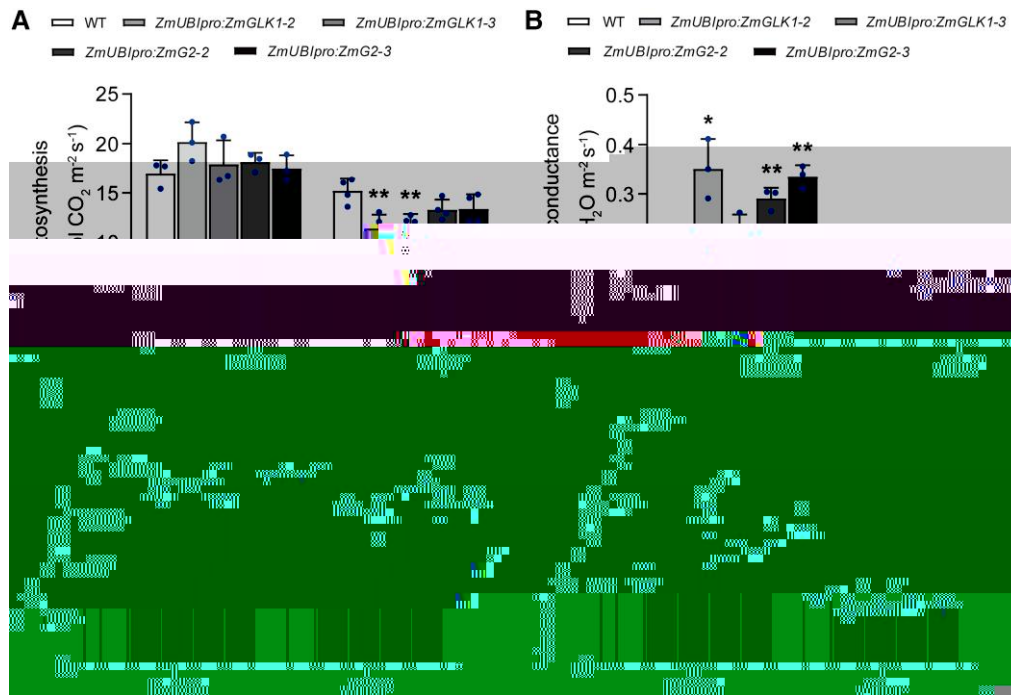
**Figure 3.** ZmGLKs conferred rapid stomatal closure to prevent water loss in rice during drought. **A)** Phenotypes of WT, *ZmUBI<sub>pro</sub>:ZmGLK1*, and *ZmUBI<sub>pro</sub>:ZmG2* rice plants during drought stress. Sixty-day-old WT, *ZmUBI<sub>pro</sub>:ZmGLK1*, and *ZmUBI<sub>pro</sub>:ZmG2* rice plants grown in soil in the greenhouse with natural light were drought stressed by withholding water for 10 d and then rewatered for a 7-d recovery period. The upper, middle, and lower panels show representative plants before drought stress, after 10 d of drought stress, and after the 7-d recovery, respectively. Scale bar: 10 cm. **B)** Survival rates of WT, *ZmUBI<sub>pro</sub>:ZmGLK1*, and *ZmUBI<sub>pro</sub>:ZmG2* rice plants after 10 d of drought stress followed by 7 d of recovery. **C)** The RWC of WT, *ZmUBI<sub>pro</sub>:ZmGLK1*, and *ZmUBI<sub>pro</sub>:ZmG2* rice leaves after 0 and 7 d of drought stress and after 7 d of recovery. **D, E)** Dynamic change of photosynthesis rate **D)** and stomatal conductance **E)** of WT, *ZmUBI<sub>pro</sub>:ZmGLK1*, and *ZmUBI<sub>pro</sub>:ZmG2* rice plants during the drought stress. Data are presented as the mean  $\pm$  SD from 3 to 6 biological replicates. \* $P < 0.05$ , \*\* $P < 0.01$  (Student's *t* test).

drought deepened, of which *ZmUBI<sub>pro</sub>:ZmGLK1* and *ZmUBI<sub>pro</sub>:ZmG2* rice plants presented lower photosynthesis rate and the stomatal conductance compared to the WT (Fig. 3, D and E). These results together clearly indicated that the rapid stomata closure was triggered by water deficiency in *ZmUBI<sub>pro</sub>:ZmGLK1* and *ZmUBI<sub>pro</sub>:ZmG2* rice plants, further contributing to the elevated drought tolerance.

### Regulation of rapid stomatal closure was ABA mediated in *ZmUBI<sub>pro</sub>:ZmGLK1* and *ZmUBI<sub>pro</sub>:ZmG2* rice plants

During the drought stress, ABA is the pivotal phytohormone that regulates stomatal movement to respond drought

(Chen et al. 2020). To further dissect the underlying mechanism associated with stomatal movement induced by *ZmGLK1* and *ZmG2*, we treated rice plants with ABA to clarify whether the rapid stomatal closure was ABA induced. After 2.5 h of applying 100  $\mu$ M ABA, *ZmUBI<sub>pro</sub>:ZmGLK1* and *ZmUBI<sub>pro</sub>:ZmG2* rice plants showed strongly decreased photosynthesis rates, accompanied with the reduced stomatal conductance (Fig. 4, A and B). Accordingly, the *C<sub>i</sub>* and transpiration rate were generally lower in *ZmUBI<sub>pro</sub>:ZmGLK1* and *ZmUBI<sub>pro</sub>:ZmG2* rice plants compared with the WT after ABA application (Fig. 4, C and D). The effects of exogenous ABA application on photosynthetic traits and stomatal conductance in



**Figure 4.** Exogenous ABA application reduced the photosynthesis rate and stomatal conductance in rice plants overexpressing *ZmGLK1* or *ZmG2* compared to the WT. **A)** Photosynthesis rates, **B)** stomatal conductance, **C)**  $C_i$  and **D)** transpiration rates of 3-wk-old WT, *ZmUBI<sub>pro</sub>:ZmGLK1*, and *ZmUBI<sub>pro</sub>:ZmG2* rice plants grown in soil before or 2.5 h after ABA treatment. Data are shown as the mean  $\pm$  SD from 3 biological replicates. \* $P < 0.05$ , \*\* $P < 0.01$  (Student's *t* test).

the WT and transgenic plants mimicked the results obtained from the drought stress treatments, which indicated the regulation of rapid stomatal closure in response to water-deficit stress conferred by *ZmGLK1* and *ZmG2* was ABA mediated.

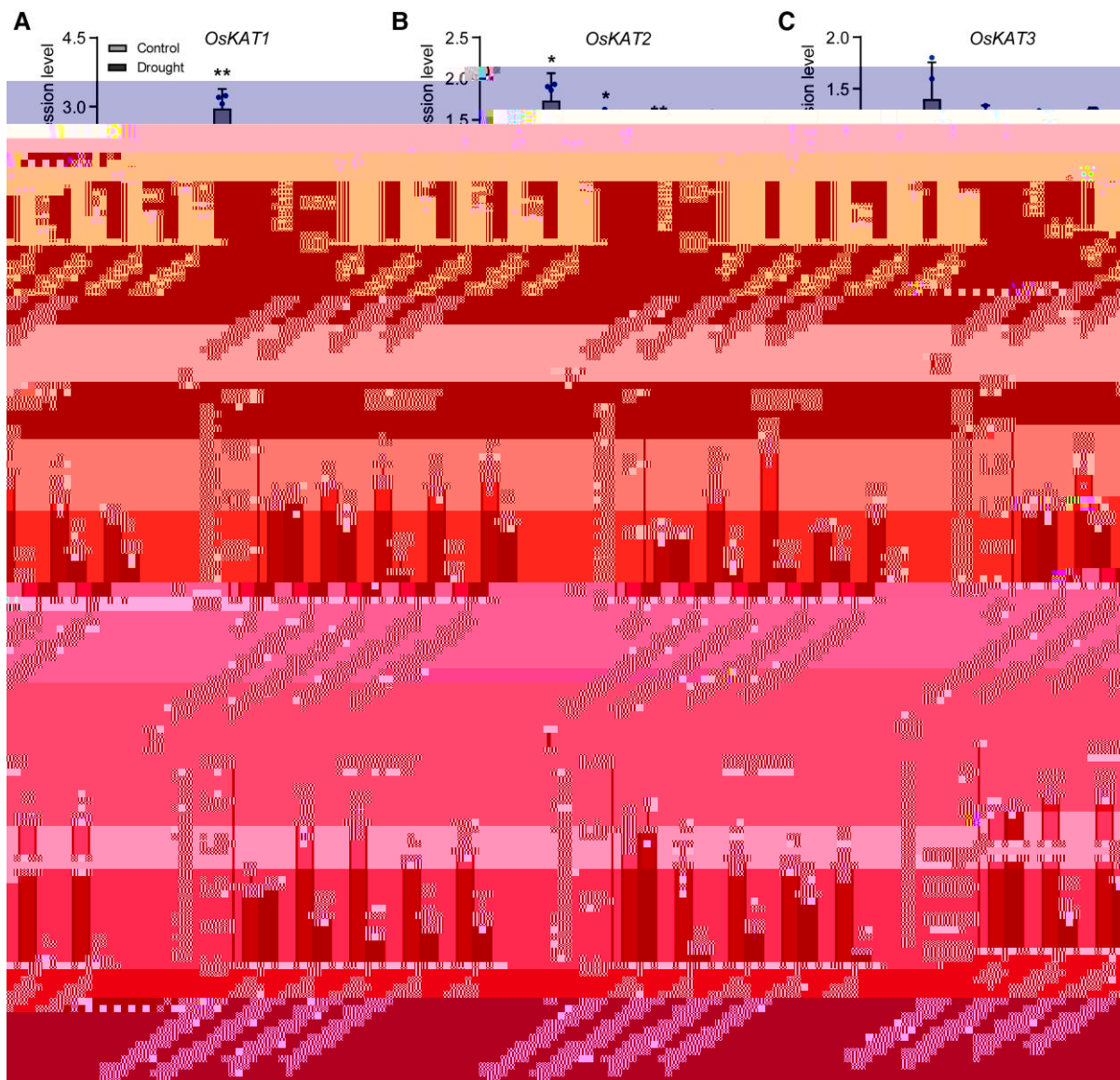
### ZmGLK1 and ZmG2 regulated stomata-related genes to promote drought tolerance

To further understand the molecular mechanisms regulated by ZmGLKs under drought stress, we next compared the expression levels of several genes associated with stomatal movement in WT, *ZmUBI<sub>pro</sub>:ZmGLK1*, and *ZmUBI<sub>pro</sub>:ZmG2* rice plants under control and drought stress conditions. Under control conditions, several key genes were highly expressed in the transgenic plants compared with the WT but profoundly downregulated in response to drought stress. These comprised 4 genes encoding proteins associated with inward rectifying shaker-like potassium channels (3 *OskATs* and 1 *OsaKT1* gene), 1  $H^+$ -ATPase (*OsaHA7*), and several stress-responsive genes (including *OsbZIP23*, *Osp5CS1*, and *OsLEA3*; Fig. 5). These results demonstrated that *ZmGLK1* and *ZmG2* improved drought tolerance by downregulating genes involved in stomatal movement when suffering from water deficit.

A genome-wide transcriptomic analysis was also conducted in WT, *ZmUBI<sub>pro</sub>:ZmGLK1*, and *ZmUBI<sub>pro</sub>:ZmG2* rice plants at 3 h after ABA treatment to investigate the global effects of *ZmGLK1* and *ZmG2* introduced by ABA, especially

on stomatal movement. WT plants clearly showed distinct expression patterns compared with *ZmUBI<sub>pro</sub>:ZmGLK1* and *ZmUBI<sub>pro</sub>:ZmG2* plants, as demonstrated by the clear separation with principal component analysis (PCA; Fig. 6A). Specifically, after ABA treatment, 702 and 775 genes were significantly upregulated in *ZmUBI<sub>pro</sub>:ZmGLK1* and *ZmUBI<sub>pro</sub>:ZmG2* plants, respectively, compared with the WT, of which 482 genes were upregulated in both transgenic lines (Fig. 6B). Gene Ontology (GO) term enrichment analysis revealed that the upregulated differentially expressed genes (DEGs) in *ZmUBI<sub>pro</sub>:ZmGLK1* and *ZmUBI<sub>pro</sub>:ZmG2* plants functioned in multiple biological processes but primarily in the ABA and water deprivation pathways (Fig. 6, C and D). Next, we performed DNA affinity purification sequencing (DAP-seq) analysis to identify genes directly regulated by the ZmGLK TFs. This analysis revealed 6,601 and 6,565 putative binding sites of *ZmGLK1* and *ZmG2* in the rice genome, respectively, with more than half of the identified sites being bound by both *ZmGLK* and *ZmG2* (Supplemental Fig. S4A). Of the 3,835 binding sites shared by *ZmGLK1* and *ZmG2*, 17.44% were localized to promoters, 8.59% to exons, and 45.26% to intergenic regions (Supplemental Fig. S4B). Motif analysis demonstrated that the most enriched core motifs found in the *ZmGLK1*- and *ZmG2*-binding regions were GCCTCT and AGATTCT (Supplemental Fig. S4, C and D). Fifty-nine genes identified from the DAP-seq data as potential targets of *ZmGLK1* and *ZmG2* in rice were also identified from the RNA-sequencing (RNA-seq) data as differentially expressed





**Figure 5.** Relative expression levels of genes involved in stomatal movement and stomatal aperture in WT, *ZmUBI<sub>pro</sub>:ZmGLK1*, and *ZmUBI<sub>pro</sub>:ZmG2* rice under normal conditions and after 7 d of drought stress. Expression levels of **A)** *OsKAT1*, **B)** *OsKAT2*, **C)** *OsKAT3*, **D)** *OsAKT1*, **E)** *OsAHA7*, **F)** *OsZIP23*, **G)** *OsLEA3*, **H)** *OsP5CS1*, and **I)** *OsERD1*. Gene expression levels were measured with RT-qPCR in the leaves of 3-wk-old rice plants grown in soil under normal conditions or drought stress for 7 d. Data are presented as the mean  $\pm$  sd from 3 biological replicates. \* $P < 0.05$ , \*\* $P < 0.01$  (Student's *t* test).

in plants overexpressing *ZmGLK1* or *ZmG2* (Fig. 6B; Supplemental Table S1). We noticed 4 upregulated DEGs were annotated to abiotic stress tolerance and showed strong binding peaks in the DAP-seq analysis simultaneously. Therefore, these genes were identified as putative target genes of *ZmGLK1* and *ZmG2* in rice, including rice genes *Filamentation Temperature Sensitive Protein H6* (*OsFtsH6*), *Cytochrome P450 Family 714 B1* (*OsCYP714B1*), *Red Chlorophyll Catabolite Reductase 1* (*OsRCCR1*), and *Subtilisin-like Protease 57* (*OsSub57*; Fig. 7, A to D). The gene expression from RNA-seq data of these 4 genes was prominently higher in *ZmUBI<sub>pro</sub>:ZmGLK1* and *ZmUBI<sub>pro</sub>:ZmG2* rice plants (Fig. 7, E to H). Further reverse transcription

quantitative PCR (RT-qPCR) analysis verified that these genes were highly induced in *ZmUBI<sub>pro</sub>:ZmGLK1* and *ZmUBI<sub>pro</sub>:ZmG2* rice under drought stress conditions (Fig. 7, I to L). These putative target genes may contribute to enhanced drought tolerance by enabling rapid stomatal movement when suffering from water deficit.

## Discussion

GLK TFs have long been regarded as some of the most important regulators of chloroplast biogenesis and photosynthetic organelle formation; they have been identified in Arabidopsis, tomato (*Solanum lycopersicum* L.), and maize



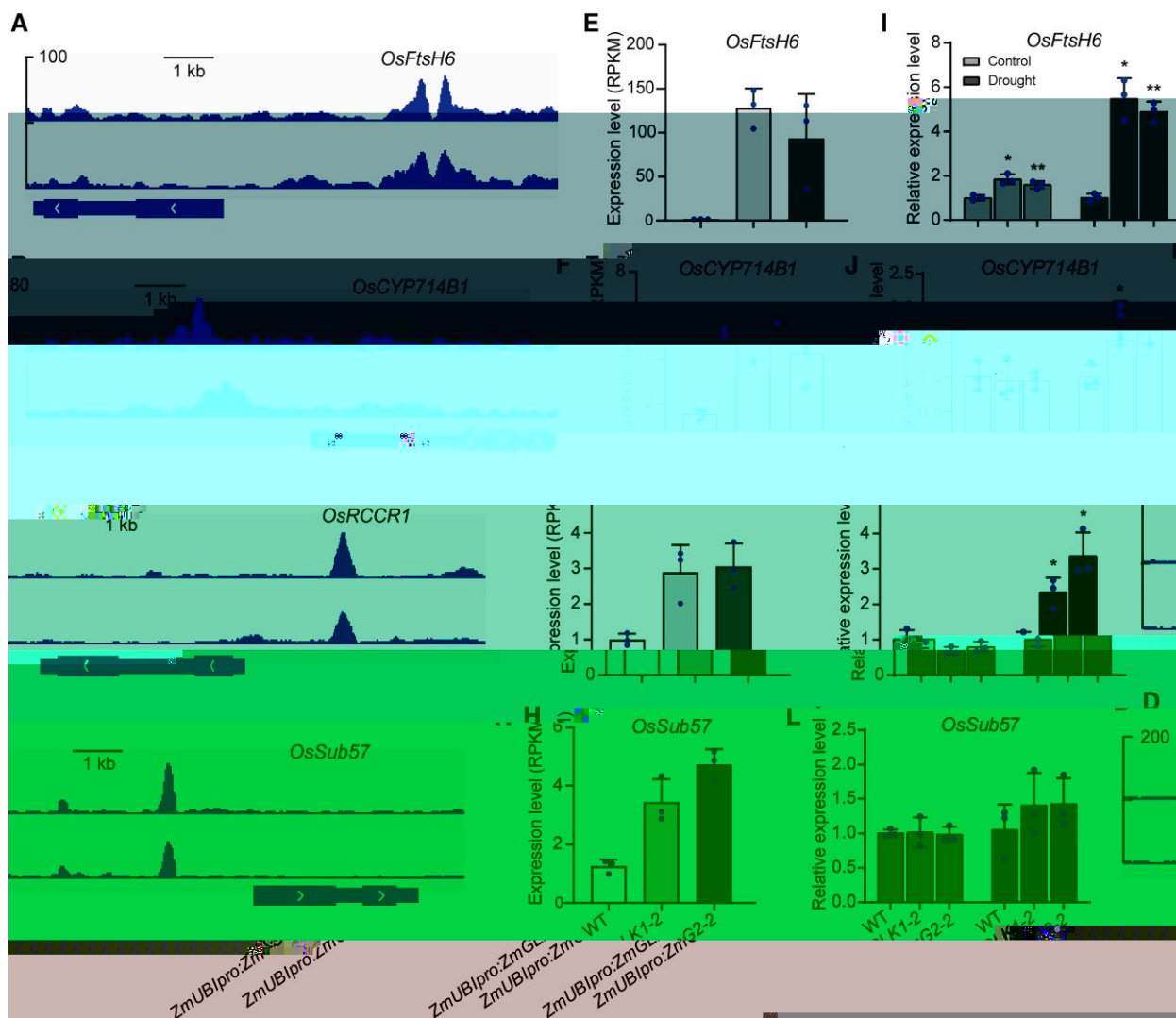
**Figure 6.** Transcriptomic analysis of WT, *ZmUBI<sub>pro</sub>:ZmGLK1*, and *ZmUBI<sub>pro</sub>:ZmG2* rice plants at 3 h after ABA treatment. **A)** PCA of gene expression in WT, *ZmUBI<sub>pro</sub>:ZmGLK1-3*, and *ZmUBI<sub>pro</sub>:ZmG2-3* rice plants based on RNA-seq data. **B)** Unique and overlapping DEGs upregulated in *ZmUBI<sub>pro</sub>:ZmGLK1* and *ZmUBI<sub>pro</sub>:ZmG2* rice plants compared to the WT and unique and overlapping *ZmGLK1* and *ZmG2* target genes identified from DAP-seq data. DEGs were identified based on  $|\log_2(\text{fold change})| > 1$  and  $P < 0.05$  by “DESeq” R package. **C, D)** GO functional categories for DEGs upregulated in *ZmUBI<sub>pro</sub>:ZmGLK1* **C)** and *ZmUBI<sub>pro</sub>:ZmG2* **D)** rice plants compared to the WT. Bubble size indicates the number of DEG counts in the corresponding GO category; bubble intensity corresponds to the  $-\log_{10}(\text{false discovery rate [FDR] value})$ ; and the x-axis indicates the ratio of DEGs in each GO category to all genes in the category.

(Rossini et al. 2001; Waters et al. 2009; Powell et al. 2012). In rice, ectopic expression of maize *GLK* genes (*ZmGLK1* and *ZmG2*) promotes a proto-Kranz status in the leaf anatomy, increasing chloroplast and mitochondrial development in rice vascular sheath cells (Wang et al. 2017). A previous study by our lab has revealed that rice plants overexpressing maize *GLK* genes have increased biomass and grain yield as a result of improved photosynthetic capacity and reduced photoinhibition under high- and fluctuating-light conditions (Li et al. 2020).

In the present study, we uncovered that overexpression of maize *GLK* genes (*ZmGLK1* and *ZmG2*) in rice enhanced drought tolerance by promoting stomatal closure. Specifically, when plants were grown under standard, well-watered conditions, we observed smaller stomatal size but higher stomatal density and stomatal aperture in rice plants overexpressing *ZmGLK1* or *ZmG2* compared with WT plants (Fig. 2, B and E). These results were consistent with earlier studies showing that *ZmGLK1* and *ZmG2* overexpression led

to increased stomatal conductance in field-grown rice (Li et al. 2020), greenhouse-grown rice (Yeh et al. 2022), and Arabidopsis (Nagatoshi et al. 2016). In contrast, under drought stress, the stomata of *ZmGLK1*- or *ZmG2*-overexpressing rice plants rapidly closed (Figs. 2B and 3E), improving drought tolerance by preventing water loss. Previous studies in rice have reported that small, high-density stomata can close quickly, thus promoting resilience against drought stress (Caine et al. 2019; Caine et al. 2023); these prior results were consistent with those of the present study. Notably, differences in stomatal status between control and drought-stressed plants as a result of *ZmGLK1* or *ZmG2* overexpression were directly caused by regulation of genes involved in stomatal movement, namely inward  $K^+$  channels and an  $H^+$ -ATPase (e.g. *OsKATs*, *OsAKT1*, and *OsAHK7*; Fig. 5). Upregulation of  $K^+$  channel genes by *ZmGLK1* or *ZmG2* overexpression under normal conditions was in line with a previous study in Arabidopsis showing that *GLK* is a positive regulator of  $K^+$  channel genes and stomatal movement (Nagatoshi et al. 2016); thus, this rapid





**Figure 7.** Putative ZmGLK1 and ZmG2 target genes in rice. **A to D)** DAP-seq indicated that ZmGLK1 and ZmG2 preferentially bound to the promoters of *OsSub57* **A)**, *OsFtsH6* **B)**, *OsCYP714B1* **C)**, and *OsRCCR1* **D)**. **E to H)** Expression levels of *OsSub57* **E)**, *OsFtsH6* **F)**, *OsCYP714B1* **G)**, and *OsRCCR1* **H)** in WT rice and in rice overexpressing *ZmGLK1* or *ZmG2* as determined with RNA-seq analysis. Gene expression was calculated in RPKM. **I to L)** Relative expression levels of *OsSub57* **I)**, *OsFtsH6* **J)**, *OsCYP714B1* **K)**, and *OsRCCR1* **L)** in WT, *ZmUBIpro:ZmGLK1*, and *ZmUBIpro:ZmG2* rice under control conditions and after 7 d of drought stress as determined with RT-qPCR. Data are presented as the mean  $\pm$  SD from 3 biological replicates. \* $P < 0.05$ , \*\* $P < 0.01$  (Student's *t* test).

stomatal closure of transgenic rice plants resulted directly from a significant reduction in the expression levels of those genes under drought conditions.

Notably, we verified that the regulation of rapid stomatal closure in response to water deficit was ABA mediated, supported by the exogenous application of ABA inducing faster stomatal closure in *ZmUBIpro:ZmGLK1* and *ZmUBIpro:ZmG2* lines compared with the WT (Fig. 4B), which mimicked the effects of drought stress. Our finding is consistent with the previous study that suggested the fast stomatal closure requires a high ABA sensitivity (Candido-Sobrinho et al. 2022). Our results also implied that ZmGLKs may function in the ABA biosynthesis pathway, as indicated by the higher ABA accumulation (Supplemental Fig. S5) along with the abundant expression

of several key genes involved in ABA biosynthesis (e.g. *OsNCED2*, *OsNCED3*, *OsAAO3*, and *OsZEP1*) in response to drought (Supplemental Fig. S6). ABA biosynthesis starts with the epoxidation of zeaxanthin, and this xanthophyll precursor therefore plays an important role in ABA biosynthesis. We previously discovered that ZmGLKs increase levels of xanthophylls, including zeaxanthin and lutein (Li et al. 2020), which may lead to the improved ABA biosynthesis in that way. Moreover, a study in *Arabidopsis* showed that GLKs directly activate the expression of *WRKY40*, and GLK-WRKY40 together negatively regulates ABA signaling (Ahmad et al. 2019), suggesting a possible regulatory role of ZmGLKs in the ABA signaling pathway. We also proposed that the  $C_4$ -like traits conferred by ZmGLKs as mentioned above may contribute to the rapid stomatal

closure. This has been demonstrated by model simulations and experimental data that major  $C_4$  crops are capable of more rapid stomatal closure compared to  $C_3$  crops in response to water deficit, resulting in the high water use efficiency (WUE) (McAusland et al. 2016; Wang et al. 2021; Ozeki et al. 2022). Notably, previous studies have demonstrated that slower stomatal closure in ferns is associated with reduced responsiveness to ABA and sugars compared to angiosperms (Lima et al. 2019; Candido-Sobrinho et al. 2022), while the rapid transport of ions and osmolytes between guard cells and subsidiary cells in grass species contributes to the fast stomatal movement (Chen et al. 2017). Rice plants overexpressing *ZmGLKs* have improved carbohydrate contents (Li et al. 2020), consistent with *SIGLK* gene expression in tomato plants (Powell et al. 2012; Nguyen et al. 2014); this may contribute to rapid stomatal closure at the metabolic level.

To further reveal the mechanism underlying *ZmGLK*-regulated stomatal movement, we conducted a comparative analysis of RNA-seq and DAP-seq data. This analysis revealed several potential target genes showing strong binding peaks, including *OsFtsH6*, *OsCYP714B1*, *OsRCCR1*, and *OsSub57* (Fig. 7). *OsFtsH6*, which belongs to the *OsFtsH* gene family, is involved in D1 turnover as part of the PSII repair cycle. D1 turnover comprises removal of damaged D1 proteins by FtsH proteases located in the chloroplast, followed by coordinated assembly of newly synthesized D1 proteins into the thylakoid membrane (Wang et al. 2016). The high levels of D1 protein observed in *ZmGLK1*- and *ZmG2*-overexpressing plants in our previous study (Li et al. 2020) prompted us to hypothesize the potential regulatory function of *ZmGLKs* on *OsFtsH6* expression. *OsCYP714B1* encodes a gibberellin (GA) 13-oxidase that plays a critical role in synthesized D1

### Stomatal trait measurements with scanning electron microscopy

Rice leaves were detached from control or drought-treated plants and immediately cut into 3 × 3 mm pieces, excluding the veins. Samples were directly fixed in 2.5% (v/v) glutaraldehyde in 0.1 M phosphate buffer (pH 7.0) and then fixed with 1% osmium tetroxide. After washing twice with 0.1 M phosphate buffer, samples were dehydrated gradually in an ethanol series (30%, 50%, 60%, 70%, 80%, 90%, and 100%) for 15 min each, followed by incubating in tertiary butanol for 35 min. Then, samples were dried using a critical point dryer, pasted on the sample stage, and then coated with gold. Stomata were observed and photographed using a SU-8010 scanning electron microscope (Hitachi, Japan). The size, number, and aperture sizes of stomata were calculated using ImageJ software.

### Quantification of endogenous ABA content

The uppermost expanded leaves of control and drought-stressed rice seedlings were detached and flash frozen in liquid nitrogen. Ground samples (100 mg each) were extracted with an acetonitrile solution containing an internal standard at 4 °C overnight. Samples were centrifuged, and the resulting supernatant was extracted again. The combined extracts were purified on a C<sub>18</sub> silica column and dried with nitrogen gas. After resolving in methanol and passing through a 0.22-μm filter, ABA was quantified on a HPLC–tandem mass spectroscopy (MS/MS) system as described by Liu et al. (2012).

### Exogenous ABA treatment

Forty-day-old rice seedlings grown in pots were sprayed with 100 μM ABA solution (containing 0.5% [v/v] Tween-20 as a surfactant) until the leaves were moist. The volume of ABA solution applied was consistent between seedlings. At 2.5 h after treatment, gas exchange parameters and stomatal traits were evaluated as described above.

### RNA extraction and RT-qPCR

The uppermost fully expanded leaves were harvested from 3-wk-old rice seedlings grown in pots under normal conditions or drought stress for 7 d. Samples were flash frozen in liquid nitrogen and ground to powder, and then total RNA was extracted with TRIzol reagent (Invitrogen). RNA purity and quantity were evaluated using a NanoDrop 2000 spectrophotometer (Thermo Fisher Scientific, USA). After DNase treatment, cDNA was synthesized from 1 μg of total RNA per sample using the RevertAid First Strand cDNA Synthesis Kit (Thermo Fisher Scientific, USA). RT-qPCR was performed using KOD SYBR Green mix with ROX (TOYOBO) on an ABI QuantStudio 6 Flex instrument (Applied Biosystems, USA). Relative transcript levels were calculated with the 2<sup>-CT</sup> method (Livak and Schmittgen 2001) with 3 biological replicates for each treatment, using *OsActin* as the internal control. Primers are listed in Supplemental Table S2.

### RNA-seq analysis

At 3 h after exogenous ABA treatment, leaves were collected from 4-wk-old rice seedlings grown in pots. Total RNA was extracted with TRIzol reagent, and then RNA integrity was assessed with the Agilent 2100 Bioanalyzer (Agilent Technologies, USA). RNA-seq libraries were constructed from WT, *ZmUBI<sub>pro</sub>:ZmGLK1-3*, and *ZmUBI<sub>pro</sub>:ZmG2-3* rice plants using the TruSeq Stranded mRNA LT Sample Prep Kit (Illumina, USA) with 3 biological replicates per line. The resulting 9 libraries were sequenced on the Illumina HiSeq X Ten sequencing platform. After removing the adaptor sequences and low-quality reads, clean reads were mapped to the *O. sativa* cv. 'Nipponbare' reference genome using HISAT (Kim et al. 2015) and Bowtie2 (Langmead et al. 2009). Gene expression levels were calculated in reads per kilobase of transcript per million mapped reads (RPKM) using Cufflinks. DEGs were identified with the "DESeq" R package. The thresholds for classification as a DEG in the transgenic lines compared to the WT were  $P < 0.05$  and  $|\log_2(\text{fold change})| > 1$ .

### DAP-seq and data analysis

The full-length coding sequences of *ZmGLK1* and *ZmG2* were amplified from cDNA of the maize accession B73. Each sequence was recombined into the pIX-HALO vector using LR Clonase II (Invitrogen). The HALO-ZmGLK1 and HALO-ZmG2 proteins were generated using 500 ng each of the pIX-HALO-ZmGLK1 and pIX-HALO-ZmG2 plasmids (Promega) following manufacturer's protocol.

ing proteins were immediately incubated with 1 μL of

Magne-HALOTag beads (Promega) for 1 h at 25 °C in 1 ×

P-40 25 °C. The be

8 times with PBST, Vj in elution buffer was

considered significant at  $P < 0.05$ . Figures were generated with GraphPad Prism 9.0 and Adobe Illustrator CS3.

### Accession numbers

Raw sequence data generated in this study have been deposited in the NCBI BioProject database under accession number PRJNA1018861 for RNA-seq and PRJNA1019016 for DAP-seq. The sequence data from this article can be found in the GenBank/EMBL data libraries under the following accession numbers: *ZmGLK1* (GenBank: AF318580) and *ZmG2* (GenBank: AF318579).

### Acknowledgments

We would like to thank Prof. Jane A. Langdale from Oxford University for kindly providing the *ZmUBI<sub>pro</sub>:ZmGLK1* and *ZmUBI<sub>pro</sub>:ZmG2* rice seeds.

### Author contributions

W.Z. and X.L. conceived and designed the experiments. X.L., J.L., S.W., Y.G., and R.G. performed most of the experiments. Z.L. and H.P. performed the DAP-seq experiment. P.W. critically commented and edited the manuscript. The manuscript was prepared by X.L., J.L., and W.Z. All authors discussed and commented on the manuscript.

### Supplemental data

The following materials are available in the online version of this article.

**Supplemental Figure S1.** Enhanced tolerance of *ZmUBI<sub>pro</sub>:ZmGLK1* and *ZmUBI<sub>pro</sub>:ZmG2* rice plants to drought stress induced by 20% PEG 6000.

**Supplemental Figure S2.** Overexpression of *ZmGLK1* or *ZmG2* in rice led to decreased stomatal conductance and photosynthetic parameters in response to drought.

**Supplemental Figure S3.** Dynamic changes of soil water content during the drought stress in the greenhouse experiment.

**Supplemental Figure S4.** Genome-wide summary of the regulatory network downstream of *ZmGLK1* and *ZmG2* based on DAP-seq data.

**Supplemental Figure S5.** Changes in endogenous ABA content in WT, *ZmUBI<sub>pro</sub>:ZmGLK1*, and *ZmUBI<sub>pro</sub>:ZmG2* rice leaves under normal conditions and after 7 d of drought stress.

**Supplemental Figure S6.** Relative expression levels of ABA biosynthesis genes in the leaves of WT, *ZmUBI<sub>pro</sub>:ZmGLK1*, and *ZmUBI<sub>pro</sub>:ZmG2* rice plants under normal conditions and after 7 d of drought stress.

**Supplemental Table S1.** Relative change of gene expression level of 59 overlapped genes from RNA-seq and DAP-seq analyses.

**Supplemental Table S2.** Primers used for RT-qPCR.

### Funding

This study was supported by grants from the National Key Research and Development Program of China

(2016YFD0300102). W.Z. was supported by the Innovation Program of the Chinese Academy of Agricultural Sciences and the Elite Youth Program of the Chinese Academy of Agricultural Sciences. X.L. was supported by the National Natural Science Foundation of China (31601237).

*Conflict of interest statement.* The authors declare that they have no conflict of interests.

### Data availability

The data underlying this article are available in the article and in its online supplementary material.

### References

- Ahmad R, Liu Y, Wang TJ, Meng Q, Yin H, Wang X, Wu Y, Nan N, Liu B, Xu ZY. GOLDEN2-LIKE transcription factors regulate *WRKY40* expression in response to abscisic acid. *Plant Physiol.* 2019;**179**(4): 1844–1860. <https://doi.org/10.1104/pp.18.01466>
- Ambavaram MM, Basu S, Krishnan A, Ramegowda V, Batlang U, Rahman L, Baisakh N, Pereira A. Coordinated regulation of photosynthesis in rice increases yield and tolerance to environmental stress. *Nat Commu.* 2014;**5**(1):5302. <https://doi.org/10.1038/ncomms6302>
- Caine RS, Harrison EL, Sloan J, Flis PM, Fischer S, Khan MS, Nguyen PT, Nguyen LT, Gray JE, Croft H. The influences of stomatal size and density on rice abiotic stress resilience. *New Phytol.* 2023;**237**(6): 2180–2195. <https://doi.org/10.1111/nph.18704>
- Caine RS, Yin XJ, Sloan J, Harrison EL, Mohammed U, Fulton T, Biswal AK, Dionora J, Chater CC, Coe RA, et al. Rice with reduced stomatal density conserves water and has improved drought tolerance under future climate conditions. *New Phytol.* 2019;**221**(1): 371–384. <https://doi.org/10.1111/nph.15344>
- Candido-Sobrinho S, Lima V, Freire F, de Souza L, Gago J, Fernie AR, Daloso DM. Metabolism-mediated mechanisms underpin the differential stomatal speediness regulation among ferns and angiosperms. *Plant, Cell Environ.* 2022;**45**(2):296–311. <https://doi.org/10.1111/pce.14232>
- Chang YM, Liu WY, Shih ACC, Shen MN, Lu CH, Lu MYJ, Yang HW, Wang TY, Chen SCC, Chen SM, et al. Characterizing regulatory and functional differentiation between maize mesophyll and bundle sheath cells by transcriptomic analysis. *Plant Physiol.* 2012;**160**(1): 165–177. <https://doi.org/10.1104/pp.112.203810>
- Chen K, Li GJ, Bressan RA, Song CP, Zhu JK, Zhao Y. Abscisic acid dynamics, signaling, and functions in plants. *J Integr Plant Biol.* 2020;**62**(1):25–54. <https://doi.org/10.1111/jipb.12899>
- Chen M, Ji M, Wen B, Liu L, Li S, Chen X, Gao D, Li L. GOLDEN 2-LIKE transcription factors of plants. *Front Plant Sci.* 2016;**7**:1509. <https://doi.org/10.3389/fpls.2016.01509>
- Chen ZH, Chen G, Dai F, Wang Y, Hills A, Ruan YL, Zhang G, Franks PJ, Nevo E, Blatt MR. Molecular evolution of grass stomata. *Trends Plant Sci.* 2017;**22**(2):124–139. <https://doi.org/10.1016/j.tplants.2016.09.005>
- FAO. Global agriculture towards 2050. In: How to feed the world in 2050, Rome. 2009.
- FAO (2021) Drought and agriculture. <https://www.fao.org/land-water/water/drought/droughtandag/en/>
- Hall LN, Rossini L, Cribb L, Langdale JA. GOLDEN 2: a novel transcriptional regulator of cellular differentiation in the maize leaf. *Plant Cell* 1998;**10**(6):925–936. <https://doi.org/10.1105/tpc.10.6.925>
- Hsu PK, Dubeaux G, Takahashi Y, Schroeder JI. Signaling mechanisms in abscisic acid-mediated stomatal closure. *Plant J.* 2021;**105**(2): 307–321. <https://doi.org/10.1111/tpj.15067>
- Kim D, Langmead B, Salzberg SL. HISAT: a fast spliced aligner with low memory requirements. *Nat Methods.* 2015;**12**(4):357–360. <https://doi.org/10.1038/nmeth.3317>

- Kim TH, Bohmer M, Hu H, Nishimura N, Schroeder JI.** Guard cell signal transduction network: advances in understanding abscisic acid, CO<sub>2</sub>, and Ca<sup>2+</sup> signaling. *Annu Rev Plant Biol.* 2010;**61**(1):561–591. <https://doi.org/10.1146/annurev-arplant-042809-112226>
- Kushiro T, Okamoto M, Nakabayashi K, Yamagishi K, Kitamura S, Asami T, Hirai N, Koshiba T, Kamiya Y, Nambara E.** The *Arabidopsis* cytochrome P450 CYP707A encodes ABA 8'-hydroxylases: key enzymes in ABA catabolism. *EMBO J.* 2004;**23**(7):1647–1656. <https://doi.org/10.1038/sj.emboj.7600121>
- Landi S, Hausman JF, Guerriero G, Esposito S.** Poaceae Vs. abiotic stress: focus on drought and salt stress, recent insights and perspectives. *Front Plant Sci.* 2017;**8**:1214. <https://doi.org/10.3389/fpls.2017.01214>
- Langmead B, Trapnell C, Pop M, Salzberg SL.** Ultrafast and memory-efficient alignment of short DNA sequences to the human genome. *Genome Biol.* 2009;**10**(3):R25. <https://doi.org/10.1186/gb-2009-10-3-r25>
- Lawson T, Viallet-Chabrand S.** Speedy stomata, photosynthesis and plant water use efficiency. *New Phytol.* 2019;**221**(1):93–98. <https://doi.org/10.1111/nph.15330>
- Li X, Wang P, Li J, Wei S, Yan Y, Yang J, Zhao M, Langdale JA, Zhou W.** Maize *GOLDEN2-LIKE* genes enhance biomass and grain yields in rice by improving photosynthesis and reducing photoinhibition. *Comm Biol.* 2020;**3**(1):151. <https://doi.org/10.1038/s42003-020-0887-3>
- Lima VF, Anjos LD, Medeiros DB, Candido-Sobrinho SA, Souza LP, Gago J, Fernie AR, Daloso DM.** The sucrose-to-malate ratio correlates with the faster CO<sub>2</sub> and light stomatal responses of angiosperms compared to ferns. *New Phytol.* 2019;**223**(4):1873–1887. <https://doi.org/10.1111/nph.15927>
- Liu H, Li X, Xiao J, Wang S.** A convenient method for simultaneous quantification of multiple phytohormones and metabolites: application in study of rice-bacterium interaction. *Plant Methods* 2012;**8**(1): 2. <https://doi.org/10.1186/1746-4811-8-2>
- Livak KJ, Schmittgen TD.** Analysis of relative gene expression data using real-time quantitative PCR and the 2<sup>-CT</sup> method. *Methods* 2001;**25**(4):402–408. <https://doi.org/10.1006/meth.2001.1262>
- Magome H, Nomura T, Hanada A, Takeda-Kamiya N, Ohnishi T, Shinma Y, Katsumata T, Kawaide H, Kamiya Y, Yamaguchi S.** *CYP714B1* and *CYP714B2* encode gibberellin 13-oxidases that reduce gibberellin activity in rice. *Proc Natl Acad Sci USA.* 2013;**110**(5): 1947–1952. <https://doi.org/10.1073/pnas.1215788110>
- Manna M, Thakur T, Chirom O, Mandlik R, Deshmukh R, Salvi P.** Transcription factors as key molecular target to strengthen the drought stress tolerance in plants. *Physiol Plant.* 2021;**172**(2): 847–868. <https://doi.org/10.1111/ppl.13268>
- McAusland L, Viallet-Chabrand S, Davey P, Baker NR, Brendel O, Lawson T.** Effects of kinetics of light-induced stomatal responses on photosynthesis and water-use efficiency. *New Phytol.* 2016;**211**(4):1209–1220. <https://doi.org/10.1111/nph.14000>
- Murmu J, Wilton M, Allard G, Pandeya R, Desveaux D, Singh J, Subramaniam R.** *Arabidopsis* GOLDEN2-LIKE (GLK) transcription factors activate jasmonic acid (JA)-dependent disease susceptibility to the biotrophic pathogen *Hyaloperonospora arabidopsidis*, as well as JA-independent plant immunity against the necrotrophic pathogen *Botrytis cinerea*. *Mol Plant Pathol.* 2014;**15**:174–184. <https://doi.org/10.1111/mp.12077>
- Nagatoshi Y, Mitsuda N, Hayashi M, Inoue S, Okuma E, Kubo A, Murata Y, Seo M, Saji H, Kinoshita T, et al.** GOLDEN 2-LIKE transcription factors for chloroplast development affect ozone tolerance through the regulation of stomatal movement. *Proc Natl Acad Sci USA.* 2016;**113**(15):4218–4223. <https://doi.org/10.1073/pnas.1513093113>
- Nguyen CV, Vrebalov JT, Gapper NE, Zheng Y, Zhong SL, Fei ZJ, Giovannoni JJ.** Tomato GOLDEN2-LIKE transcription factors reveal molecular gradients that function during fruit development and ripening. *Plant Cell* 2014;**26**(2):585–601. <https://doi.org/10.1105/tpc.113.118794>
- Ozeki K, Miyazawa Y, Sugiura D.** Rapid stomatal closure contributes to higher water use efficiency in major C<sub>4</sub> compared to C<sub>3</sub> Poaceae crops. *Plant Physiol.* 2022;**189**(1):188–203. <https://doi.org/10.1093/plphys/kiac040>
- Pandey S, Zhang W, Assmann SM.** Roles of ion channels and transporters in guard cell signal transduction. *FEBS Lett.* 2007;**581**(12): 2325–2336. <https://doi.org/10.1016/j.febslet.2007.04.008>
- Powell AL, Nguyen CV, Hill T, Cheng KL, Figueroa-Balderas R, Aktas H, Ashrafi H, Pons C, Fernandez-Munoz R, Vicente A, et al.** Uniform ripening encodes a *Golden 2-like* transcription factor regulating tomato fruit chloroplast development. *Science* 2012;**336**(6089): 1711–1715. <https://doi.org/10.1126/science.1222218>
- Ray DK, Mueller ND, West PC, Foley JA.** Yield trends are insufficient to double global crop production by 2050. *PLoS One* 2013;**8**(6):e66428. <https://doi.org/10.1371/journal.pone.0066428>
- Ray DK, Ramankutty N, Mueller ND, West PC, Foley JA.** Recent patterns of crop yield growth and stagnation. *Nat Commun.* 2012;**3**(1): 1293. <https://doi.org/10.1038/ncomms2296>
- Rossini L, Cribb L, Martin DJ, Langdale JA.** The maize *Golden2* gene defines a novel class of transcriptional regulators in plants. *Plant Cell* 2001;**13**(5):1231–1244. <https://doi.org/10.1105/tpc.13.5.1231>
- Seo M, Koshiba T.** Complex regulation of ABA biosynthesis in plants. *Trends Plant Sci.* 2002;**7**(1):41–48. [https://doi.org/10.1016/S1360-1385\(01\)02187-2](https://doi.org/10.1016/S1360-1385(01)02187-2)
- Sierla M, Hörak H, Overmyer K, Waszczak C, Yarmolinsky D, Maierhofer T, Vainonen JP, Salojärvi J, Denessiouk K, Laanemets K, et al.** The receptor-like pseudokinase GHR1 is required for stomatal closure. *Plant Cell* 2018;**30**(11):2813–2837. <https://doi.org/10.1105/tpc.18.00441>
- Tang Y, Li M, Chen Y, Wu P, Wu G, Jiang H.** Knockdown of *OsPAO* and *OsRCCR1* cause different plant death phenotypes in rice. *J Plant Physiol.* 2011;**168**(16):1952–1959. <https://doi.org/10.1016/j.jplph.2011.05.026>
- Tilman D, Balzer C, Hill J, Befort BL.** Global food demand and the sustainable intensification of agriculture. *Proc Natl Acad Sci USA.* 2011;**108**(50):20260–20264. <https://doi.org/10.1073/pnas.1116437108>
- Todaka D, Nakashima K, Shinozaki K, Yamaguchi-Shinozaki K.** Toward understanding transcriptional regulatory networks in abiotic stress responses and tolerance in rice. *Rice* 2012;**5**(1):6. <https://doi.org/10.1186/1939-8433-5-6>
- Wang F, Liu J, Chen j nyw@DZXZ,raTdnjChen ngcKIKTj nf-kQ,T,Tdnj n KTj nTwZNN,**